Kertas Ulasan/Review Articles

The Role of Reactive Oxygen Species in Pressure-Dependent Myogenic Tone (Peranan Spesis Oksigen Reaktif dalam Tonus Miogenik yang Bergantung dengan Tekanan)

SATIRAH ZAINALABIDIN, PAUL COATS & ROGER M. WADSWORTH

ABSTRACT

Myogenic tone is the response of the vascular smooth muscle to an increase in intraluminal pressure with vasoconstriction and with vasodilation when the pressure is decreased. Such myogenic tone contributes a level of physiological basal tone in response to neurohumoral stimuli. In spite of myogenic tone discovery by Sir William Bayliss 100 years ago, questions still remain regarding the underlying signaling mechanism of the myogenic response. Studies have shown that increased intraluminal pressure or wall tension leads to membrane depolarization, voltage-operated calcium channel (VOCC), stretch-activated cation (SAC) channels, extracelullar matrix (ECM) and actin cytoskeleton. Recently, evidence has shown a potential role for reactive oxygen species (ROS) as a key signalling mediator in the genesis of myogenic tone. The identification of the primary mechanosensors in the initiation of pressure-dependent myogenic tone is essential as these components could be potential therapeutical targets in the future.

Keywords: Microcirculation, ROS, myogenic tone

ABSTRAK

Tonus miogenik adalah respon vasokonstriksi otot licin vaskular ke atas peningkatan tekanan intralumina dan respon vasodilatasi pula ke atas penurunan tekanan intralumina. Kompensasi tonus miogenik menyumbang kepada pengawalaturan tonus basal dalam keadaan fisiologi terhadap rangsangan neurohormon. Walaupun Sir William Bayliss telah menemui tonus miogenik pada 100 tahun yang lalu, namun mekanisme signal di sebalik respons miogenik masih belum dikenal pasti secara mendalam. Kajian lepas telah menunjukkan bahawa peningkatan tekanan intralumina ataupun ketegangan dinding vaskular menyebabkan depolarisasi membran, rangsangan kanal kalsium voltan-tergantung, kation ketegangan-terangsang, matrik ekstraselular dan sitoskeleton aktin. Bukti telah menunjukkan potensi sepsis oksigen reaktif (ROS) sebagai perantara signal semasa mekanisme tonus miogenik. Pengenalpastian dan pemahaman mengenai mekanosensor primary yang merangsang permulaan tonus miogenik bergantung-tekanan adalah penting sebagai potensi terapeutik di masa hadapan.

Kata kunci: Mikrosirkulasi, ROS, tonus miogenik

AUTOREGULATION IN RESISTANCE ARTERIES

In the cardiovascular system, blood vessel function is related to its structure. Specifically, major arteries of the trunk are the elastic conductance arteries with large diameter and low resistance. On the contrary, distal arteries are muscular arteries of small diameter and high resistance that are critical for blood flow or pressure within the microcirculation system. Under physiological conditions, the viscoelasticity of the large arteries results in pulsatile pressure and flow so that the microvasculature can mediate blood supply steadily. The microvessels proximal to the arterioles with lumen diameter less than 200 µm were defined by Mulvany and Aalkjaer (1990) as the vessels offering the greatest resistance. The resistance value can vary depending for the entire vascular system, for the vascular network in selected organs and for individual vessels (Zweifach 1991), therefore, the size of resistance arteries may vary and be more than 200 µm diameter.

Resistance arteries have the ability to alter their diameter independent of neural and hormonal control, in order to keep a relatively constant blood flow. The efficiency of autoregulation varies from tissue to tissue with those organs most vital for survival, e.g. brain, heart, kidney, demonstrating the most marked autoregulatory capacity. A number of local intrinsic control mechanisms have been proposed to account for the homeostatic properties of autoregulation, which are the combination of metabolites such as lactate and adenosine, flow-dependent regulation and myogenic tone (Hill et al. 2006; Henrion 2005).

Pressure-dependent myogenic tone in small resistance arteries is defined as the intrinsic property of vascular smooth muscle (VSM) cells to respond to changes in transmural pressure. Sir William Bayliss was the first to discover myogenic tone in 1902 when he recorded large increases in the volume of a dog's hind limb following the release of brief aortic occlusion (Bayliss 1902). Bayliss considered the blood volume increase response too rapid to be mediated by accumulation of metabolites and concluded that a significant component of vascular tone could be modulated by changes in intravascular pressure. Bjorn Folkow (1949) supported Bayliss's theory demonstrating that denervated preparations of blood vessels developed pressure-dependent vascular tone and autoregulation of blood flow was neurohormonal-independent (Folkow 1949). However, in spite of more than 100 years of myogenic tone study, the knowledge of the exact primary sensors to the signalling mechanisms that allow changes of intraluminal pressure to be converted accordingly into appropriate arterial diameter, remain incomplete and controversial. This has been a key challenge and a true understanding of the underlying mechanism is important in order to tackle the treatment of the pathological states.

PRESSURE-DEPENDENT MYOGENIC TONE MECHANISMS

Myogenic tone shows an inverse relationship between pressure and diameter, where an increase of intraluminal pressure causes a decrease in diameter, and vice versa. Moreover, the strength of the myogenic response varies depending on the diameter and the particular vascular bed. In the hamster cheek pouch, relative myogenic responsiveness increased with decreasing vessel size in second- and third-order arterioles, whereas fourth-order arterioles were substantially less responsive than thirdorder arterioles (Davis 1993). Relative myogenic tone varies between different vascular beds, with rat cerebral (Gokina et al. 2005) and skeletal muscle vessels (Coats 2010) exhibiting more prominent myogenic response than small mesenteric vessels (Chin et al. 2007). Differences in the strength of the myogenic response between vessels with similar diameter have also been observed within the network of a vascular bed. For example, there was a weak myogenic response in subendocardial but a strong myogenic response in subepicardial porcine arteries (Kuo et al. 1988).

According to the reviews by Hill et al. (2006) and Khavandi et al. (2009), myogenic behaviour can be divided into three phases. The first consists of myogenic or basal tone development, associated with a large increase in L-type VOCC-mediated membrane depolarization and $[Ca^{2+}]_{i}$. The second phase is known as myogenic reactivity, where further constriction happens in response to an increase in intraluminal pressure. The [Ca²⁺], concentration is maintained at this phase, but [Ca²⁺], sensitization of the mechanical apparatus is thought to occur. The third phase is called forced dilation, when the wall of the artery is unable to maintain a constriction against mounting pressure. There are a number of candidates that could be mediating myogenic response, however there is no agreement on whether it is the vessel wall tension (Davis et al. 1992, Dunn & Gardiner 1997, Brekke et al. 2002), stretch-activated cation (SAC) channels (Davis et al.

1992), VOCC (McCarron et al. 1997), extracelullar matrix (ECM) (Davis et al. 2001) or actin cytoskeleton (Cipolla et al. 2002). Knowledge about the cellular mechanisms underlying myogenic tone is important and could be exploited for therapeutic intervention, as myogenic responsiveness is commonly altered in vascular diseases. Therefore, it is crucial to understand the stimuli, sensor, signal transduction pathways involving second messengers, regulation of contractile proteins and finally the adjustment for vasomotor tone.

ION CHANNELS

Putative plasma membrane ion channels functioning as primary mechanosensors in mediating arteriolar myogenic vasoconstriction include VOCC (Kotecha & Hill 2005; Knot & Nelson 1998), SAC (Luchessi et al. 2004), TRPC (Mederos y Schnitzler et al. 2008), large conductance Ca²⁺-activated K⁺ channel (BK_{Ca}) (Dong et al. 2009; Cheranov & Jaggar 2006), voltage-gated potassium channel (K_v) (Sobey et al. 1998) and Ca²⁺-activated chloride channels (Cl_{Ca}) (Dimitropoulou et al. 2001).

A plethora of evidence has shown that the pressuredependent myogenic response is mediated by an increase of membrane depolarization and [Ca2+], in VSM, accompanied by the classical Ca²⁺-calmodulin-induced myosin light chain (MLC) phosphorylation (Harder 1984; Davis & Hill 1999; Hill et al. 2001; Schubert et al. 2008). The pioneering study by Harder (1984) was the first to show that an increase in pressure caused depolarization and therefore an increase in Ca²⁺ influx and vasoconstriction, within the cerebral artery. A subsequent study showed that the increased Ca²⁺ influx occurred in parallel with the decrease in diameter (Knot & Nelson 1998). Incubation in a Ca²⁺-free solution or the administration of the Ca²⁺-entry blocker diltiazem, led to a complete loss of myogenic tone (Bevan 1982). The Ca²⁺ influx has been shown to depend primarily on L-type VOCC which can be inhibited by nifedipine (Kotecha & Hill 2005; Coats et al. 2001; Scotland et al. 2004; VanBavel et al. 1998).

The primary events that lead to an increase of [Ca²⁺]_i are the release of Ca²⁺ from intracellular stores and transmembrane influx. However, a study found the internal ryanodine-sensitive Ca2+ store to be insignificant in myogenic constriction, indicating that CICR is a minor contributor to the response (McCarron et al. 1997). Recently, extracellular Ca²⁺ has been reviewed by Smajilovic and Tfelt-Hansen (2007) for eliciting the role as a first messenger through a G-protein coupled receptor (GPCR), namely the Ca²⁺-sensing receptor (CaR). In subcutaneous arteries, the CaR was found to modulate pressure-dependent myogenic tone, while PKC was acting as its negative regulator (Ohanian et al. 2005). However, the CaR paradigm is relatively new in the area of myogenic tone and more investigation is needed to clarify its function. Another recent study that supports GCPR as a

mechanosensor is where $G_{q'11}$ -coupled receptor, a GCPR, was activated and stimulated β -arrestin recruitment in response to increased pressure (Mederos y Schnitzler et al. 2008). Subsequently, this process lead to TRPC-mediated membrane depolarization and enhanced myogenic constriction in renal and cerebral arteries.

Although Ca^{2+} has been shown as a necessary key signalling component in myogenic constriction, some studies have shown that only a minimal rise in $[Ca^{2+}]_i$ is required to elicit myogenic constriction (D'Angelo et al. 1997; VanBavel et al. 1998). Therefore, Ca^{2+} is not the only determinant of steady-state constriction and there are Ca^{2+} -independent regulatory systems involving protein kinase C (PKC), Rho/Rho kinase, protein tyrosine kinase and cytoskeleton, that accompany myogenic constriction (Lagaud et al. 2002; Gokina et al. 2005; Massett et al. 2002; Jarajapu & Knot 2002).

INTRACELLULAR SECOND MESSENGERS

There has been a growing body of evidence that shows Ca²⁺ sensitivity regulation integrates with Ca²⁺-independent mechanisms in the myogenic constriction, via intracellular second messengers signalling in VSM. Pressure or stretch acts as the stimulus for G proteins, which then stimulate the membrane enzyme phospholipase C (PLC) that specifically hydrolyzes phosphatidylinositol-4,5-biphosphate (PIP,). PIP, is split into two second messengers, diacylglycerol (DAG) and inotisol-1,4,5-triphosphate (IP₃) (Narayanan et al. 1994). Both IP, and DAG have been shown to increase with the level of intraluminal pressure and to remain elevated for the duration of the pressure increase (Narayanan et al. 1994). The increase in the level of intracellular IP,, induced by stretch, may be responsible for part of the pressureinduced increase of [Ca²⁺]; (Jarajapu & Knot 2002). The increase in DAG, which is an activator of protein kinase C (PKC), implicates the participation of PKC in the myogenic response (Gokina et al. 1999; Wesselman et al. 2001). Indeed, the PKC activator, phorbol-12,13-dibutyrate (PDBu) was found to increase the level of Ca2+, and this effect was abolished by nifedipine, an L-type Ca²⁺ channel blocker (Lin et al. 1998; Ohanian et al. 2005). This suggests that the PKC-enhancing effect by PDBu was through the activation of L-type Ca²⁺ channels and this could increase Ca²⁺ sensitivity resulting in myogenic constriction (Lin et al. 1998; Gokina et al. 1999). Consistent with the fact that PKC phosphorylates mitogen-activated protein kinase (MAPK), it has been shown that inhibition of MAPK reduced the Ca2+ sensitivity and hence, attenuated the pressure-dependent myogenic tone (Massett et al. 2002; Lagaud et al. 1999). Another potential pathway regulating myogenic tone is the RhoA/Rho kinase signaling pathway. Results have shown that RhoA/Rho kinase pathway to be activated by mechanical stretch, leading to an increase in intracellular [Ca²⁺] sensitivity. This could potentially inhibit myosin phosphatase and/or activate actin cytoskeleton polymerization, followed by increased myogenic tone (Gokina et al. 2005; Jarajapu & Knot 2005). Another interesting key candidate in modulating myogenic tone is the epidermal growth factor receptor (EGFR). A study in mesenteric arteries has shown pressure/stretch to stimulate metalloproteinases 2/9 (MMP-2/9), leading to endogenous HB-EGF (heparin-binding EGF-like growth factor) release and EGFR transactivation (Lucchesi et al. 2004). This mechanism was specific to myogenic tone as contractions to ANG II and KCl were independent of EGFR transactivation. The underlying mechanism within the VSM was not investigated, but it was speculated that MMP-2/9 activation may involve integrins (Belmadani et al. 2008) or SACs (Scotland et al. 2004), and the downstream signalling of EGFR activation may be PKC-dependent (Jarajapu & Knot 2005) or through Nox/ROS (Csanyi et al. 2009).

CYTOCHROME P-450

The cytochrome P-450 (CYP) enzymes metabolize endogenous arachidonic acid (AA) and are referred to as the third pathway of AA metabolism, the others being cyclooxygenases and lipooxygenases. The role of CYP has been reviewed as a candidate mediator in signaling events culminating in pressure-dependent myogenic tone in renal, skeletal muscle and pulmonary arteries (Terashvili et al. 2006; Frisbee et al. 2001; Kaley 2000; Parker et al. 2005). Phospholipase A_{2} (PLA₂) is an enzyme which hydrolyzes phospholipids and produces AA. This AA can be further metabolized to produce autocrine vasoconstrictor 20-HETE (20-hydroxyeicosatetraenoic acid) and vasodilator 11,12-EET (11,12-epoxyeicosatrienoic acid), which could enhance ROS release. Stimulation of VSM cells by stretch, pressure and flow could trigger CYP to release 20-HETE, which inhibits the opening of Ca^{2+} -activated K⁺ (K_c) channels in VSM, causing depolarization, and eventually increases Ca²⁺ entry and promotes myogenic constriction (Frisbee et al. 2001). As for 11,12-EET, it exhibits biological activities opposite to 20-HETE; where it vasodilates blood vessels and stimulates hyperpolarization via K_{Ca} channels in VSM cells (Imig 1999). Hence, the dual counterbalancing activity of these two AA metabolites may present a mechanism for balanced regulation of myogenic tone regulation.

REACTIVE OXYGEN SPECIES

Reactive oxygen species and oxidative stress have been synonymous in pathological conditions for decades. Recently, a growing body of evidence has shown that ROS could play a role in regulating pressure-dependent myogenic tone (Lecarpentier 2007; Keller et al. 2006). Besides ROS, another emerging new candidate is the actin cytoskeleton (Faraci 2006). Figure 1 summarizes the overall putative mechanotransduction pathways of pressure-dependent myogenic tone regulation.



FIGURE 1. Overview of possible pathways involved in the myogenic response. The possible pathways may involve ion channels, intracellular second messengers, cytochrome P450 and the newly arised new candidates, ROS and actin cytoskeleton. PIP₂ (phosphatidylinositol (4,5)-bisphosphate); PLC (phospholipase C); IP₃ (inositol trisphosphate); DAG (diacylglycerol); PKC (protein kinase C); VOCC (voltage-operated calcium channel); PLA₂ (phospholipase A₂); 20-HETE (20-hydroxyeicosatetraenoic acid); KCa (calcium-activated potassium channel); Ca²⁺ (calcium); CaM (calmodulin); MLCK (myosin light-chain kinase); myosin-P (myosin-phosphate); SR (sarcoplamic reticulum)

REACTIVE OXYGEN SPECIES IN CARDIOVASCULAR HEALTH AND DISEASES

REACTIVE OXYGEN SPECIES AND OXIDATIVE STRESS

ROS are classically considered as a toxic by-product of metabolism in oxidative stress (Taniyama & Griendling 2003; Dworakowski et al. 2006). When ROS is produced excessively, overwhelming of endogenous antioxidant systems and/or during downregulation of antioxidant systems, oxidative stress occurs (Griendling & Fitzgerald 2003; Taniyama & Griendling 2003; Dworakowski et al. 2006). Dysregulation due to oxidative stress has been shown to contribute substantially to the progression of cardiovascular diseases such as hypertension (Schiffrin 2004; Intengan et al. 1999), diabetes (Sachidanandam et al. 2007), hypercholesterolaemia (Ishikawa et al. 2004), cardiac hypertrophy (Lang 2002) and myocardial ischaemia (Ferrari et al. 2004).

The term 'oxidative stress' may be an oversimplification as it is used to cover the diverse and complex role of ROS in physiological and pathophysiological conditions. Over the past decade, studies have shown that ROS at relatively low cellular levels may influence metabolism in physiological conditions (Lecarpentier 2007; Keller et al. 2006) and may be involved with the normal regulation of vascular tone and structure (Faraci 2006). ROS may have direct and indirect effects on VSM, and there are studies suggesting that both relaxation and contraction of vascular muscle may occur, depending on the tissue model and the physiological circumstances.

METABOLISM OF ROS

ROS includes free radicals such as superoxide anion (O_{2}) and hydroxyl radical (OH-), and nonradical species such as hydrogen peroxide (H₂O₂) (Hamilton et al. 2004). Superoxide dismutase (SOD) metabolizes O₂⁻ to H₂O₂. There are three isoforms for SOD: CuZn-containing cytosolic SOD1, Mn-containing SOD2 and CuZn-containing extracellular SOD3. Peroxidases such as catalase and glutathione peroxidase further metabolize H₂O₂ to O₂ and water. H₂O₂ can also be transformed to OH⁻ and O₂⁻ to peroxynitrite (ONOO⁻) (Faraci 2006; Miller et al. 2006; Hamilton et al. 2004). Almost all types of vascular cell produce O₂⁻ and H₂O₂ (Lyle & Griendling 2005). Primarily, ROS within a blood vessel is produced by the membranebound enzyme Nox (Griendling et al. 1994), but ROS has been shown to also be produced from adventitial fibroblasts (Pagano et al. 1997), cyclooxygenase (Sobey et al. 1998;

Miller et al. 2006) as well as eNOS (Matoba et al. 2000). There are many stimuli which activate Nox, including ANG II (Mehta and Griendling, 2007), ET-1 (An et al. 2007), shear stress (Hwang et al. 2003), tumor necrosis factor- α (TNF- α) (Al-Mehdi and Fisher 1998), metabolic factors including hyperglycaemia, insulin (Dworakowski et al. 2006), hypercholesterolemia (Ishikawa et al. 2004), and vascular endothelial growth factor (VEGF) (Abid et al. 2007).

 O_2^- is highly reactive, but it has a very short half-life and poor cell permeability. Therefore, the impact of $O_2^$ on vascular function could be instant but short-lasting. The reaction of O_2^- with EDNO is very rapid, 3 times faster than the dismutation of O_2^- , resulting in reduced NO bioavailability and increased ONOO⁻. Conversely, H_2O_2 is relatively stable and highly diffusible between cells. Therefore, H_2O_2 is regarded as one of the most important ROS molecules for modulating vascular function. It has been reported as an EDHF in the mesenteric artery (Matoba et al. 2000) and partially mediates flow-dependent vasodilatation in coronary arterioles (Miura et al. 2003). Figure 2 shows the general metabolism of ROS and the associated complications in cardiovascular diseases.



FIGURE 2. Generation of O_2^- and H_2O_2 from O_2 in vascular tissue and the associated complications in cardiovascular diseases. GSSG, oxidized glutathione; GSH, reduced glutathione; SOD, superoxide dismutase; ONOO-, peroxynitrite; NO, nitric oxide; VSM, vascular smooth muscle; EC, endothelial cells

NADPH OXIDASE SUBUNITS

The Nox in the vasculature consists of p22^{phox}, p47^{phox} and novel homologues of gp91^{phox}, which are NOX1, NOX2 and NOX4 (Seshiah et al. 2002; Cave 2008). These isoforms are different between cell types and they have separate locations within the cell. The evidence is becoming increasingly clear that the individual Nox isoforms have delineated roles within the cell and are linked with specific downstream effects (Cave et al. 2005; Cave 2008).

ANG II AS AN NADPH OXIDASE/ ROS ACTIVATOR

ANG II stimulates Nox and ROS activity in all cells within the vascular wall (Griendling & Ushio-Fukai 2000; Pedruzzi et al. 2004; Gonzalez et al. 2008). ANG II not only stimulates Nox-dependent O₂⁻ formation in VSM cells but also in cardiac, endothelial, mesangial cells and adventitial fibroblasts (Zhang et al. 1999; Pedruzzi et al. 2004; Mehta & Griendling 2007). In rabbit aortic adventitial fibroblasts, induction of Nox subunit p67phox expression was caused by ANG II-induced O₂⁻ (Pagano et al. 1998). In rat VSM, subunit p22^{phox} has been shown to be a component in the ANG II-induced hypertrophy (Ushio-Fukai et al. 1996). A member of a new family of gp91^{phox}, NOX1, showed a role in mediating ANG II-induced O₂⁻ formation via redox signalling in VSM cells (Lassegue et al. 2001). Angiotensin AT, receptors (AT,R) are believed to be mainly located in the VSM cells and to induce vasoconstriction (Maeso et al. 1996; Touyz & Schiffrin 2000), while angiotensin AT, receptors (AT,R) mediate endothelium-dependent vasodilation (Carey et al. 2000).

In addition to stimulating Nox to release ROS, ANG II has also been found to stimulate various downstream signalling pathways in relation to pressure-dependent myogenic tone. For instance, intraluminal pressure facilitated by ANG II may activate AT, R which in turn causes transactivation of the platelet-derived growth factor (PDGF), then through the Ras-Raf pathway activates extracellular signal-regulated kinase 1/2 (ERK1/2) (Matrougui et al. 2000, Eskildsen-Helmond & Mulvany 2003). However, how the mechanical loading can cause such acute and rapid activation is still unknown. Suggested pathways may include the PLC-IP, pathway resulting in increase of $[Ca^{2+}]$ and Ca^{2+} -dependent ERK1/2 stimulation, or the PLC-DAG pathway whereby PKC can activate ERK1/2 (Eskildsen-Helmond & Mulvany 2003). Other interesting supporting findings were that AT₁R stimulation could initiate G-to-F actin and strengthen the cytoskeletal structure (Martinez-Lemus 2008) or it could activate TRPC-mediated membrane depolarization and cause myogenic tone (Mederos y Schnitzler et al. 2008).

ROS IN THE BIOLOGY OF VASCULAR TONE

The first direct evidence that endogenous ROS can act as a signalling molecule was found by Kontos et al. (1984). Their study found that dilation by topical sodium arachidonate and bradykinin was inhibited by catalase and SOD in cat cerebral artery. Thus, O_2^- and H_2O_2 or radical derivatives such as OH⁻ were likely to be the mediators from sodium arachidonate and bradykinin that caused vasodilation. Since then, a growing body of evidence has demonstrated ROS as potent intracellular and intercellular second messengers to modulate many downstream signalling molecules via $[Ca^{2+}]_i$ -dependent and $[Ca^{2+}]_i$ -independent pathways and eventually cause substantial interplay in myogenic response.

The major effect of O_2^- was thought to be peripheral vasocontraction by its reaction with EDNO, as shown widely in oxidative stress. It is now known that in cerebral arteries, O_{2}^{-} can cause both constriction and dilation (Wei et al. 1996; Didion & Faraci 2002). The K⁺ channel blocker, tetraethylammonium (TEA), reduced cerebral relaxation to O_2^{-} , suggesting that K_{C_a} channels are involved (Wei et al. 1996). H₂O₂ has also been shown to induce both vasorelaxation and vasoconstriction, depending on the vascular bed and the pre-constrictor agent. H₂O₂ from the endothelium may act as a vasodilatory EDHF in murine (Kimura et al. 2002), human mesenteric arteries (Matoba et al. 2002) and human coronary arterioles (Miura et al. 2003). At higher concentration (>1 mM), H_2O_2 has been found to cause vasoconstriction followed by vasodilatation (Miller et al. 2005). Dilatation of rat cerebral arterioles in response to exogenous H₂O₂, or endogenous bradykininderived H₂O₂ is mediated by activation of K_{Ca} channels (Sobey et al. 1997). In other studies, mitochondrial-derived ROS dilated cerebral arteries by increasing K_{Ca} channel activity via Ca²⁺ sparks (Xi et al. 2005), whereas H₂O₂ dilated cerebral arterioles by activating ATP-sensitive K⁺ channels (Wei et al. 1996). Thus, activation of K⁺ channels may be a major mechanism of dilatation in response to H₂O₂ in the microcirculation. In a study on canine basilar arteries, different types of exogenous ROS elicit different characteristics of contraction, whereby H2O2 showed the fast onset and a transient contraction, O_2^{-} showed slow onset and a transient contraction, and OH- showed a sustained contraction (Tosaka et al. 2002). This shows the importance of identifying the types of radical and nonradical species in the pathological mechanisms as each ROS could play a different role.

ROS

From the study of ROS in cultured cells to vascular tone, the interest of its role in modulating pressure-dependent myogenic tone has been growing. Exogenous H_2O_2 was found to increase myogenic tone in tail arterioles (Nowicki et al. 2001) and mesenteric arteries (Lucchesi et al. 2005). In addition, endogenous ROS (H_2O_2) was produced from Nox after dipheleneiodonium (DPI) and N-acetylcysteine (NAC) treatment, and mice lacking rac1 and p47^{phox} showed inhibited myogenic tone (Nowicki et al. 2001). Another supporting study showed myogenic tone was dependent on mitochondria-derived ROS (H_2O_2) during cold (28°C), which stimulated RhoA/Rho kinase signalling and smooth muscle α_{2c} -adrenoceptors, a mechanism that could be relevant to Raynaud's pathophysiology (Bailey et al. 2005). Although H₂O₂ caused constriction in the study, other studies showed that it could also act as a vasodilator (EDHF) (Miura et al. 2003; Matoba et al. 2002; Drouin et al. 2007). The dual effects of H2O2 were also observed in a study by Luchessi et al. (2005) where low concentrations of exogenous H₂O₂ caused vasodilation when the K⁺ channel was compromised by KCl, and the opposite effect was seen when KCl was removed in pressurized mesenteric arteries. H₂O₂ also exhibits a biphasic effect, causing vasoconstriction at low concentrations and vasodilation at high concentrations, also possibly mediated via K⁺ channels (Cseko et al. 2004). With these observations, it could be possible that in pathophysiological conditions where K⁺ channel activity is altered, a transition from the physiological vasodilatory effect of H₂O₂ to a contractile effect may play an important role in increasing peripheral resistance and pathogenesis of hypertension (Cseko et al. 2004). In hamster gracilis muscle, Sk1/S1P was shown to mediate pressure-dependent myogenic tone by stimulating the Rac/Nox/O₂⁻ pathway (Keller et al. 2006). Inhibition of a specific Nox subunit was shown to reduce pressure-dependent myogenic tone, but it did not affect the $[Ca^{2+}]_{i}$, which suggested that ROS is playing a modulatory rather than obligatory role in regulation of Ca²⁺ sensitivity (Keller et al. 2006). This observation is very important as Sk1/S1P could be a potential therapeutic target, although there should be a balance for the beneficial formation of O_2^{-} as a physiological signalling molecule.

The role of ROS has also been investigated in diseased animal models. NOX2-derived O₂⁻ may contribute to the enhanced myogenic response in renal afferent arterioles in spontaneously hypertensive rats (SHR), but not in normal rats (Ren et al. 2010). According to this study, the role of ROS is complex, because on one hand, increased O₂⁻ in afferent arterioles could add to hypertension development. But on the other hand, after hypertension is established, O₂-induced myogenic enhancement could provide protection from glomerular barotrauma and injury in SHRs. In streptozotocin-induced diabetic rats, cerebral myogenic tone was increased in parallel with decreased large conductance BK_{Ca} channel activity and increased H₂O₂ (Dong et al. 2009). Rotenone, an inhibitor of the mitochondrial electron transport chain complex I, partially reversed the myogenic response, H₂O₂ levels and BK_{C_a} effect. This indicates that BK_{C_a} channels may be related to the inhibitory effects of rotenone on ROS (H_2O_2) production and the myogenic tone of diabetic rats (Dong et al. 2009).

The important role of ROS as a signalling molecule in pressure-dependent myogenic tone is indisputable. However, there arises a significant dilemma in determining the borderline of the beneficial and the hazardous effect of ROS. Additionally, ROS members are highly diverse and able to elicit multiple effects within the same tissue. Paravicini & Sobey (2003) have highlighted the importance of the delicate balance between O_2^- and H_2O_2 mediated by SOD when studying vascular oxidative stress. Therefore, it is essential to understand the exact mechanism categorized in order to determine how ROS can be 'good' or 'bad' signalling molecules.

POTENTIAL THERAPEUTIC IMPLICATIONS OF ARTERIAL MYOGENIC TONE

Considering our current knowledge of myogenic tone regulation, an ability to adjust vascular resistance through specific modulation of myogenic mechanism would enable normal neurohormonal mechanisms to interact with the therapeutically treated tone. Therefore, identifying the primary mechanosensors in the initiation of pressuredependent myogenic tone is essential as these components could be targets for drug development. For example, in situations in which the cardiovascular system is depressed in shock and low-flow states, a drug increasing myogenic tone would be beneficial. On the contrary, in hyperdynamic conditions, reduction of drug-induced myogenic tone would be needed.

The possibility of the adventitia, especially the adventitial fibroblast, being a potential platform for intravascular local drug delivery is very intriguing. Work in the future should focus on understanding ways to control adventitial fibroblast activation, to reduce unwanted changes in vascular structure. As the adventitial fibroblasts contain Nox subunits, the development of specific inhibitors of Nox would provide pharmacological tools to completely elucidate the roles of these isoforms in vascular physiology and pharmacology, particularly, in pressure-dependent myogenic tone. To date, there is only one such compound i.e. plumbagin that has been reported to inhibit Nox/O₂⁻ production by NOX4 in cultured kidney embryonic cells (Ding et al. 2005). However, this compound was found to be carcinogenic and whether this compound selectively inhibits NOX4 activity or acts merely as a ROS scavenger was not investigated. Rey et al. (2001) designed a Nox inhibitor known as gp91ds-tat, which competitively binds to p47^{phox} and prevents the assembly of the enzyme. To date, this peptide has been said to be the most successful Nox inhibitor because of its selectivity. However, it is uncertain if gp91ds-tat inhibits other Noxcontaining isoforms of Nox, such as NOX1 or NOX4. In conclusion, it is essential to understand the specific Nox subunit source of ROS within the adventitial fibroblasts as it may become a potential therapeutic target in treating vascular diseases.

REFERENCES

Abid, M. R., Spokes, K. C., Shih, S. C. & Aird, W. C. 2007. NADPH oxidase activity selectively modulates vascular endothelial growth factor signaling pathways. *J Biol Chem*, 282: 35373-85.

- Al-Mehdi, A.B. & Fisher, A.B. 1998. Invited editorial on "tumor necrosis factor-alpha in ischemia and reperfusion injury in rat lungs". J. Appl. Physiol. 85: 2003-4.
- An, S. J., Boyd, R., Zhu, M., Chapman, A., Pimentel, D. R. & Wang, H. D. 2007. NADPH oxidase mediates angiotensin II-induced endothelin-1 expression in vascular adventitial fibroblasts. *Cardiovasc Res*, 75: 702-9.
- Bailey, S.R., Mitra, S., Flavahan, S. & Flavahan, N.A. 2005. Reactive oxygen species from smooth muscle mitochondria initiate cold-induced constriction of cutaneous arteries. *Am. J. Physiol. Heart Circ. Physiol.* 289: H243-50.
- Bayliss, W.M. 1902. On the local reactions of the arterial wall to changes of internal pressure. *J. Physiol.* 28: 220-31.
- Belmadani, S., Zerfaoui, M., Boulares, H.A., Palen, D.I. & Matrougui, K. 2008. Microvessel vascular smooth muscle cells contribute to collagen type I deposition through ERK1/2 MAP kinase, alphavbeta3-integrin, and TGF-beta1 in response to ANG II and high glucose. *Am. J. Physiol. Heart Circ. Physiol.* 295: H69-76.
- Bevan, J.A. 1982. Selective action of diltiazem on cerebral vascular smooth muscle in the rabbit: antagonism of extrinsic but not intrinsic maintained tone. *Am. J. Cardiol.* 49: 519-24.
- Brekke, J.F., Gokina, N.I. & Osol, G. 2002. Vascular smooth muscle cell stress as a determinant of cerebral artery myogenic tone. *Am. J. Physiol. Heart Circ. Physiol.* 283: H2210-6.
- Carey, R.M., Wang, Z.Q. & Siragy, H.M. 2000. Update: role of the angiotensin type-2 (AT(2)) receptor in blood pressure regulation. *Curr. Hypertens Rep.* 2: 198-201.
- Cave, A. 2008. Selective targeting of NADPH oxidase for cardiovascular protection. *Curr Opin Pharmacol.*, 9: 208-13.
- Cave, A., Grieve, D., Johar, S., Zhang, M. & Shah, A. M. 2005. NADPH oxidase-derived reactive oxygen species in cardiac pathophysiology. *Philos Trans R Soc Lond B Biol Sci*, 360: 2327-34.
- Cheranov, S. Y. & Jaggar, J. H. 2006. TNF-alpha dilates cerebral arteries via NAD(P)H oxidase-dependent Ca²⁺ spark activation. *Am J Physiol Cell Physiol*, 290: C964-71.
- Chin, L.C., Achike, F.I. & Mustafa, M.R. 2007. Hydrogen peroxide modulates angiotensin II-induced contraction of mesenteric arteries from streptozotocin-induced diabetic rats. *Vascul Pharmacol.* 46: 223-8.
- Cipolla, M.J., Gokina, N.I. & Osol, G. 2002. Pressure-induced actin polymerization in vascular smooth muscle as a mechanism underlying myogenic behavior. *FASEB J.* 16: 72-76.
- Coats, P. 2010. The effect of peripheral vascular disease on structure and function of resistance arteries isolated from human skeletal muscle. *Clin. Physiol. Funct. Imaging.* 30: 192-7.
- Coats, P., Johnston, F., Macdonald, J., Mcmurray, J.J. & Hillier, C. 2001. Signalling mechanisms underlying the myogenic response in human subcutaneous resistance arteries. *Cardiovasc Res.* 49: 828-37.
- Csanyi, G., Taylor, W.R. & Pagano, P.J. 2009. NOX and inflammation in the vascular adventitia. *Free Radic. Biol. Med.* 47: 1254-66.
- Cseko, C., Bagi, Z. & Koller, A. 2004. Biphasic effect of hydrogen peroxide on skeletal muscle arteriolar tone via activation of endothelial and smooth muscle signaling pathways. *J. Appl. Physiol.* 97: 1130-7.

- D'angelo, G., Davis, M.J. & Meininger, G.A. 1997. Calcium and mechanotransduction of the myogenic response. Am. J. Physiol. 273: H175-82.
- Davis, M.J. 1993. Myogenic response gradient in an arteriolar network. Am J Physiol, 264: H2168-79.
- Davis, M.J., Donovitz, J.A. & Hood, J.D. 1992. Stretch-activated single-channel and whole cell currents in vascular smooth muscle cells. *Am. J. Physiol.* 262: C1083-8.
- Davis, M.J. & Hill, M.A. 1999. Signaling mechanisms underlying the vascular myogenic response. *Physiol Rev.* 79: 387-423.
- Davis, M.J., Wu, X., Nurkiewicz, T.R., Kawasaki, J., Davis, G.E., Hill, M.A. & Meininger, G.A. 2001. Integrins and mechanotransduction of the vascular myogenic response. *Am. J. Physiol. Heart. Circ Physiol.* 280: H1427-33.
- Didion, S. P. & Faraci, F. M. 2002. Effects of NADH and NADPH on superoxide levels and cerebral vascular tone. *Am J Physiol Heart Circ Physiol*, 282: H688-95.
- Dimitropoulou, C., White, R.E., Fuchs, L., Zhang, H., Catravas, J.D. & Carrier, G.O. 2001. Angiotensin II relaxes microvessels via the AT(2) receptor and Ca(2+)-activated K(+) (BK(Ca)) channels. *Hypertension*. 37: 301-7.
- Ding, Y., Chen, Z.J., Liu, S., Che, D., Vetter, M. & Chang, C.H. 2005. Inhibition of Nox-4 activity by plumbagin, a plantderived bioactive naphthoquinone. *J. Pharm. Pharmacol.* 57: 111-6.
- Dong, L., Xie, M.J., Zhang, P., JI, L.L., Liu, W.C., Dong, M.Q. & Gao, F. 2009. Rotenone partially reverses decreased BK Ca currents in cerebral artery smooth muscle cells from streptozotocin-induced diabetic mice. *Clin. Exp. Pharmacol. Physiol.* 36: 57-64.
- Drouin, A., Thorin-Trescases, N., Hamel, E., Falck, J.R. & Thorin, E. 2007. Endothelial nitric oxide synthase activation leads to dilatory H₂O₂ production in mouse cerebral arteries. *Cardiovasc Res.* 73: 73-81.
- Dunn, W.R. & Gardiner, S.M. 1997. Differential alteration in vascular structure of resistance arteries isolated from the cerebral and mesenteric vascular beds of transgenic [(mRen-2)27], hypertensive rats. *Hypertension*. 29: 1140-7.
- Dworakowski, R., Anilkumar, N., Zhang, M. & Shah, A.M. 2006. Redox signalling involving NADPH oxidase-derived reactive oxygen species. *Biochem Soc Trans.* 34: 960-4.
- Eskildsen-Helmond, Y.E. & Mulvany, M.J. 2003. Pressureinduced activation of extracellular signal-regulated kinase 1/2 in small arteries. *Hypertension*, 41: 891-7.
- Faraci, F.M. 2006. Reactive oxygen species: influence on cerebral vascular tone. J. Appl. Physiol. 100: 739-43.
- Ferrari, R., Guardigli, G., Mele, D., Percoco, G.F., Ceconi, C. & Curello, S. 2004. Oxidative stress during myocardial ischaemia and heart failure. *Curr. Pharm Des.* 10: 1699-711.
- Folkow, B. 1949. Intravascular pressure as a factor regulating the tone of the small vessels. *Acta Physiol Scand*, 17: 289-310.
- Frisbee, J.C., Roman, R.J., Falck, J.R., Krishna, U.M. & Lombard, J.H. 2001. 20-HETE contributes to myogenic activation of skeletal muscle resistance arteries in Brown Norway and Sprague-Dawley rats. *Microcirculation*. 8: 45-55.
- Gokina, N.I., Knot, H.J., Nelson, M.T. & Osol, G. 1999. Increased Ca²⁺ sensitivity as a key mechanism of PKC-induced constriction in pressurized cerebral arteries. *Am J. Physiol.* 277: H1178-88.

- Gokina, N.I., Park, K.M., Mcelroy-Yaggy, K. & Osol, G. 2005. Effects of Rho kinase inhibition on cerebral artery myogenic tone and reactivity. J. Appl. Physiol. 98: 1940-8.
- Gonzalez, J.M., Somoza, B., Conde, M.V., Fernandez-Alfonso, M.S., Gonzalez, M.C. & ARRIBAS, S.M. 2008. Hypertension increases middle cerebral artery resting tone in spontaneously hypertensive rats: role of tonic vasoactive factor availability. *Clin Sci (Lond)* 114: 651-9.
- Griendling, K.K., Minieri, C.A., Ollerenshaw, J.D. & Alexander, R.W. 1994. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res.* 74: 1141-8.
- Griendling, K.K. & FitzGerald, G.A. 2003. Oxidative stress and cardiovascular injury: Part I: basic mechanisms and in vivo monitoring of ROS. *Circulation* 108: 1912-6.
- Griendling, K.K. & Ushio-Fukai, M. 2000. Reactive oxygen species as mediators of angiotensin II signaling. *Regul. Pept.* 91: 21-7.
- Hamilton, C.A., Miller, W.H., Al-Benna, S., Brosnan, M.J., Drummond, R.D., Mcbride, M.W. & Dominiczak, A.F. 2004. Strategies to reduce oxidative stress in cardiovascular disease. *Clin. Sci. (Lond)* 106: 219-34.
- Harder, D.R. 1984. Pressure-dependent membrane depolarization in cat middle cerebral artery. Circ Res. 55: 197-202.
- Henrion, D. 2005. Pressure and flow-dependent tone in resistance arteries. Role of myogenic tone. Arch. Mal. Coeur. Vaiss 98: 913-21.
- Hill, M.A., Davis, M.J., Meininger, G.A., Potocnik, S.J. & Murphy, T.V. 2006. Arteriolar myogenic signalling mechanisms: Implications for local vascular function. *Clin Hemorheol. Microcirc.* 34: 67-79.
- Hill, M.A., Zou, H., Potocnik, S.J., Meininger, G.A. & Davis, M.J. 2001. Invited review: arteriolar smooth muscle mechanotransduction: Ca(2+) signaling pathways underlying myogenic reactivity. J. Appl. Physiol. 91: 973-83.
- Hwang, J., Saha, A., Boo, Y. C., Sorescu, G. P., McNally, J. S., Holland, S. M., Dikalov, S., Giddens, D. P., Griendling, K. K., Harrison, D. G. & Jo, H. 2003. Oscillatory shear stress stimulates endothelial production of O²⁻ from p47phoxdependent NAD(P)H oxidases, leading to monocyte adhesion. *J Biol Chem* 278: 47291-8.
- Imig, J.D. 1999. Epoxyeicosatrienoic acids. Biosynthesis, regulation, and actions. *Methods Mol. Biol.* 120: 173-92.
- Intengan, H.D., Deng, L.Y., Li, J.S. & Schiffrin, E.L. 1999. Mechanics and composition of human subcutaneous resistance arteries in essential hypertension. *Hypertension* 33: 569-74.
- Ishikawa, M., Stokes, K.Y., Zhang, J.H., Nanda, A. & Granger, D.N. 2004. Cerebral microvascular responses to hypercholesterolemia: roles of NADPH oxidase and P-selectin. *Circ. Res.* 94: 239-44.
- Jarajapu, Y.P. & Knot, H.J. 2002. Role of phospholipase C in development of myogenic tone in rat posterior cerebral arteries. Am. J. Physiol. Heart Circ. Physiol. 283: H2234-8.
- Jarajapu, Y.P. & Knot, H.J. 2005. Relative contribution of Rho kinase and protein kinase C to myogenic tone in rat cerebral arteries in hypertension. *Am. J. Physiol. Heart Circ. Physiol.* 289: H1917-22.
- Kaley, G. 2000. Regulation of vascular tone: role of 20-HETE in the modulation of myogenic reactivity. *Circ. Res.* 87: 4-5.

- Keller, M., Lidington, D., Vogel, L., Peter, B.F., Sohn, H.-Y., Pagano, P.J., Pitson, S., Spiegel, S., Pohl, U. & Bolz, S.-S. 2006. Sphingosine kinase functionally links elevated transmural pressure and increased reactive oxygen species formation in resistance arteries. *FASEB J*: 05-4075fje.
- Khavandi, K., Greenstein, A.S., Sonoyama, K., Withers, S., Price, A., Malik, R.A. & Heagerty, A.M. 2009. Myogenic tone and small artery remodelling: insight into diabetic nephropathy. *Nephrol Dial Transplant* 24: 361-9.
- Kimura, K., Tsuda, K., Moriwaki, C., Kawabe, T., Hamada, M., Obana, M., Baba, A., Hano, T. & Nishio, I. 2002. Leukemia inhibitory factor relaxes arteries through endotheliumdependent mechanism. *Biochem Biophys Res Commun.* 294: 359-62.
- Knot, H.J. & Nelson, M.T. 1998. Regulation of arterial diameter and wall [Ca²⁺] in cerebral arteries of rat by membrane potential and intravascular pressure. *J. Physiol.* 508(1): 199-209.
- Kontos, H.A., Wei, E.P., Povlishock, J.T. & Christman, C.W. 1984. Oxygen radicals mediate the cerebral arteriolar dilation from arachidonate and bradykinin in cats. *Circ Res.* 55: 295-303.
- Kotecha, N. & Hill, M.A. 2005. Myogenic contraction in rat skeletal muscle arterioles: smooth muscle membrane potential and Ca(2+) signaling. *Am. J. Physiol. Heart Circ.* Physiol. 289: H1326-34.
- Kuo, L., Davis, M.J. & Chilian, W.M. 1988. Myogenic activity in isolated subepicardial and subendocardial coronary arterioles. Am. J. Physiol. 255: H1558-62.
- Lagaud, G., Gaudreault, N., Moore, E.D., Van Breemen, C. & Laher, I. 2002. Pressure-dependent myogenic constriction of cerebral arteries occurs independently of voltagedependent activation. Am. J. Physiol. Heart Circ. Physiol. 283: H2187-95.
- Lagaud, G.J., Lam, E., Lui, A., Van Breemen, C. & Laher, I. 1999. Nonspecific inhibition of myogenic tone by PD98059, a MEK1 inhibitor, in rat middle cerebral arteries. *Biochem. Biophys. Res. Commun.* 257: 523-7.
- Lang, D. 2002. Cardiac hypertrophy and oxidative stress: a leap of faith or stark reality? *Heart*. 87: 316-7.
- Lassegue, B., Sorescu, D., Szocs, K., Yin, Q., Akers, M., Zhang, Y., Grant, S.L., Lambeth, J.D. & Griendling, K.K. 2001. Novel gp91(phox) homologues in vascular smooth muscle cells: nox1 mediates angiotensin II-induced superoxide formation and redox-sensitive signaling pathways. *Circ Res.* 88: 888-94.
- Lecarpentier, Y. 2007. Physiological role of free radicals in skeletal muscles. *J. Appl. Physiol.* 103: 1917-8.
- Lin, M.J., Liu, S.H. & Lin-Shiau, S.Y. 1998. Phorbol esterinduced contractions of mouse detrusor muscle are inhibited by nifedipine. *Naunyn Schmiedebergs Arch. Pharmacol.* 357: 553-7.
- Lucchesi, P.A., Belmadani, S. & Matrougui, K. 2005. Hydrogen peroxide acts as both vasodilator and vasoconstrictor in the control of perfused mouse mesenteric resistance arteries. *J. Hypertens.* 23: 571-9.
- Lucchesi, P.A., Sabri, A., Belmadani, S. & Matrougui, K. 2004. Involvement of metalloproteinases 2/9 in epidermal growth factor receptor transactivation in pressure-induced myogenic tone in mouse mesenteric resistance arteries. *Circulation*. 110: 3587-93.

- Lyle, A.N. & Griendling, K.K. 2006. Modulation of vascular smooth muscle signaling by reactive oxygen species. *Physiology (Bethesda)* 21: 269-80.
- Maeso, R., Navarro-Cid, J., Munoz-Garcia, R., Rodrigo, E., Ruilope, L.M., Lahera, V. & Cachofeiro, V. 1996. Losartan reduces phenylephrine constrictor response in aortic rings from spontaneously hypertensive rats. Role of nitric oxide and angiotensin II type 2 receptors. *Hypertension* 28: 967-72.
- Martinez-Lemus, L. A. 2008. Persistent agonist-induced vasoconstriction is not required for angiotensin II to mediate inward remodeling of isolated arterioles with myogenic tone. *J. Vasc. Res.* 45: 211-21.
- Massett, M.P., Ungvari, Z., Csiszar, A., Kaley, G. & Koller, A. 2002. Different roles of PKC and MAP kinases in arteriolar constrictions to pressure and agonists. *Am. J. Physiol. Heart Circ. Physiol.* 283: H2282-7.
- Matoba, T., Shimokawa, H., Kubota, H., Morikawa, K., Fujiki, T., Kunihiro, I., Mukai, Y., Hirakawa, Y. & Takeshita, A. 2002. Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in human mesenteric arteries. *Biochem. Biophys. Res. Commun.* 290: 909-13.
- Matoba, T., Shimokawa, H., Nakashima, M., Hirakawa, Y., Mukai, Y., Hirano, K., Kanaide, H. & Takeshita, A. 2000. Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in mice. J. Clin. Invest. 106: 1521-30.
- Matrougui, K., Eskildsen-Helmond, Y.E., Fiebeler, A., Henrion, D., Levy, B.I., Tedgui, A. & Mulvany, M.J. 2000. Angiotensin II stimulates extracellular signal-regulated kinase activity in intact pressurized rat mesenteric resistance arteries. *Hypertension*. 36: 617-21.
- Mccarron, J.G., Crichton, C.A., Langton, P.D., Mackenzie, A. & Smith, G.L. 1997. Myogenic contraction by modulation of voltage-dependent calcium currents in isolated rat cerebral arteries. J. Physiol. 498(2): 371-9.
- Mederos Y Schnitzler, M., Storch, U., Meibers, S., Nurwakagari, P., Breit, A., Essin, K., Gollasch, M. & Gudermann, T. 2008. Gq-coupled receptors as mechanosensors mediating myogenic vasoconstriction. *Embo. J.* 27: 3092-103.
- Mehta, P.K. & Griendling, K.K. 2007. Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. *Am. J. Physiol. Cell Physiol.* 292: C82-97.
- Miller, A.A., Drummond, G.R., Schmidt, H.H. & Sobey, C.G. 2005. NADPH oxidase activity and function are profoundly greater in cerebral versus systemic arteries. *Circ. Res.* 97: 1055-62.
- Miller, A.A., Drummond, G.R. & Sobey, C.G. 2006. Reactive oxygen species in the cerebral circulation: are they all bad? *Antioxid Redox Signal* 8: 1113-20.
- Miura, H., Bosnjak, J.J., Ning, G., Saito, T., Miura, M. & Gutterman, D.D. 2003. Role for hydrogen peroxide in flow-induced dilation of human coronary arterioles. *Circ. Res.* 92: e31-40.
- Mulvany, M.J. & Aalkjaer, C. 1990. Structure and function of small arteries. *Physiol Rev.* 70: 921-61.
- Narayanan, J., Imig, M., Roman, R.J. & Harder, D.R. 1994. Pressurization of isolated renal arteries increases inositol trisphosphate and diacylglycerol. *Am. J. Physiol.* 266: H1840-5.
- Nowicki, P.T., Flavahan, S., Hassanain, H., Mitra, S., Holland, S., Goldschmidt-Clermont, P.J. & Flavahan, N.A. 2001.

Redox signaling of the arteriolar myogenic response. *Circ. Res.* 89: 114-6.

- Ohanian, J., Gatfield, K.M., Ward, D.T. & Ohanian, V. 2005. Evidence for a functional calcium-sensing receptor that modulates myogenic tone in rat subcutaneous small arteries. *Am. J. Physiol. Heart Circ. Physiol.* 288: H1756-62.
- Pagano, P.J., Chanock, S.J., Siwik, D.A., Colucci, W.S. & Clark, J.K. 1998. Angiotensin II induces p67phox mRNA expression and NADPH oxidase superoxide generation in rabbit aortic adventitial fibroblasts. *Hypertension* 32: 331-7.
- Pagano, P.J., Clark, J.K., Cifuentes-Pagano, M.E., Clark, S.M., Callis, G.M. & Quinn, M.T. 1997. Localization of a constitutively active, phagocyte-like NADPH oxidase in rabbit aortic adventitia: enhancement by angiotensin II. *Proc. Natl. Acad. Sci. U.S.A.* 94: 14483-8.
- Paravicini, T.M. & Sobey, C.G. 2003. Cerebral vascular effects of reactive oxygen species: recent evidence for a role of NADPH-oxidase. *Clin Exp Pharmacol Physiol.* 30: 855-9.
- Parker, T.A., Grover, T.R., Kinsella, J.P., Falck, J.R. & Abman, S.H. 2005. Inhibition of 20-HETE abolishes the myogenic response during NOS antagonism in the ovine fetal pulmonary circulation. *Am. J. Physiol. Lung Cell Mol. Physiol.* 289: L261-7.
- Pedruzzi, E., Guichard, C., Ollivier, V., Driss, F., Fay, M., Prunet, C., Marie, J.C., Pouzet, C., Samadi, M., Elbim, C., O'dowd, Y., Bens, M., Vandewalle, A., Gougerot-Pocidalo, M.A., Lizard, G. & Ogier-Denis, E. 2004. NAD(P)H oxidase Nox-4 mediates 7-ketocholesterol-induced endoplasmic reticulum stress and apoptosis in human aortic smooth muscle cells. *Mol. Cell Biol.* 24: 10703-17.
- Ren, Y., D'ambrosio, M.A., Liu, R., Pagano, P.J., Garvin, J.L. & Carretero, O.A. 2010. Enhanced myogenic response in the afferent arteriole of spontaneously hypertensive rats. *Am. J. Physiol. Heart Circ. Physiol.* 298: H1769-75.
- Rey, F.E., Cifuentes, M.E., Kiarash, A., Quinn, M.T. & Pagano, P.J. 2001. Novel competitive inhibitor of NAD(P)H oxidase assembly attenuates vascular O₂⁻ and systolic blood pressure in mice. *Circ Res.* 89: 408-14.
- Sachidanandam, K., Portik-Dobos, V., Harris, A.K., Hutchinson, J.R., Muller, E., Johnson, M.H. & Ergul, A. 2007. Evidence for vasculoprotective effects of ETB receptors in resistance artery remodeling in diabetes. *Diabetes* 56: 2753-8.
- Schiffrin, E.L. 2004. Remodeling of resistance arteries in essential hypertension and effects of antihypertensive treatment. *Am J Hypertens* 17: 1192-200.
- Schubert, R., Lidington, D. & Bolz, S.S. 2008. The emerging role of Ca²⁺ sensitivity regulation in promoting myogenic vasoconstriction. *Cardiovasc Res.* 77: 8-18.
- Scotland, R.S., Chauhan, S., Davis, C., De Felipe, C., Hunt, S., Kabir, J., Kotsonis, P., Oh, U. & Ahluwalia, A. 2004. Vanilloid receptor TRPV1, sensory C-fibers, and vascular autoregulation: a novel mechanism involved in myogenic constriction. *Circ. Res.* 95: 1027-34.
- Seshiah, P.N., Weber, D.S., Rocic, P., Valppu, L., Taniyama, Y. & Griendling, K.K. 2002. Angiotensin II stimulation of NAD(P)H oxidase activity: upstream mediators. *Circ Res.* 91: 406-13.
- Smajilovic, S. & Tfelt-Hansen, J. 2007. Calcium acts as a first messenger through the calcium-sensing receptor in the cardiovascular system. *Cardiovasc Res.* 75: 457-67.

- Sobey, C.G., Heistad, D.D. & Faraci, F.M. 1997. Mechanisms of bradykinin-induced cerebral vasodilatation in rats. Evidence that reactive oxygen species activate K+ channels. *Stroke*. 28: 2290-4.
- Sobey, C.G., Heistad, D.D. & Faraci, F.M. 1998. Potassium channels mediate dilatation of cerebral arterioles in response to arachidonate. *Am. J. Physiol.* 275: H1606-12.
- Taniyama, Y. & Griendling, K.K. 2003. Reactive oxygen species in the vasculature: molecular and cellular mechanisms. *Hypertension* 42: 1075-81.
- Terashvili, M., Pratt, P.F., Gebremedhin, D., Narayanan, J. & Harder, D.R. 2006. Reactive oxygen species cerebral autoregulation in health and disease. *Pediatr. Clin. North. Am*, 53: 1029-37.
- Tosaka, M., Hashiba, Y., Saito, N., Imai, H., Shimizu, T. & Sasaki, T. 2002. Contractile responses to reactive oxygen species in the canine basilar artery in vitro: selective inhibitory effect of MCI-186, a new hydroxyl radical scavenger. *Acta Neurochir* (*Wien*), 144: 1305-10; discussion 1310.
- Touyz, R.M. & Schiffrin, E.L. 2000. Signal transduction mechanisms mediating the physiological and pathophysiological actions of angiotensin II in vascular smooth muscle cells. *Pharmacol Rev.* 52: 639-72.
- Ushio-Fukai, M., Zafari, A.M., Fukui, T., Ishizaka, N. & Griendling, K.K. 1996. p22phox is a critical component of the superoxide-generating NADH/NADPH oxidase system

Satirah Zainalabidin Biomedical Science Programme School of Diagnostic & Applied Health Sciences Faculty of Health Sciences Universiti Kebangsaan Malaysia Jalan Raja Muda Abdul Aziz 50300 Kuala Lumpur, MALAYSIA

Correspondence author: Satirah Zainalabidin Email address: satirah@fskb.ukm.my Tel: 603-92897684, Fax: 603-91456635

Received: January 2012 Accepted for publication: February 2012 and regulates angiotensin II-induced hypertrophy in vascular smooth muscle cells. *J Biol Chem.* 271: 23317-21.

- Vanbavel, E., Wesselman, J.P. & Spaan, J.A. 1998. Myogenic activation and calcium sensitivity of cannulated rat mesenteric small arteries. *Circ. Res.* 82: 210-20.
- Wei, E.P., Kontos, H.A. & Beckman, J.S. 1996. Mechanisms of cerebral vasodilation by superoxide, hydrogen peroxide, and peroxynitrite. *Am. J. Physiol.* 271: H1262-6.
- Wesselman, J.P., Spaan, J.A., Van Der Meulen, E.T. & Vanbavel, E. 2001. Role of protein kinase C in myogenic calciumcontraction coupling of rat cannulated mesenteric small arteries. *Clin. Exp. Pharmacol. Physiol.* 28: 848-55.
- Xi, Q., Cheranov, S.Y. & Jaggar, J.H. 2005. Mitochondria-derived reactive oxygen species dilate cerebral arteries by activating Ca²⁺ sparks. *Circ. Res.* 97: 354-62.
- Zhang, H., Schmeisser, A., Garlichs, C.D., Plotze, K., Damme, U., Mugge, A. & Daniel, W.G. 1999. Angiotensin II-induced superoxide anion generation in human vascular endothelial cells: role of membrane-bound NADH-/NADPH-oxidases. *Cardiovasc Res.* 44: 215-22.
- Zweifach, B.W. 1991. Vascular Resistance: Structural vs functional basis. In Bevan, J.A., Halpern, W., Mulvany, M.J. (Eds), *The Resistance Vasculature*. New Jersey: Humana Pres.

Paul Coats Roger M. Wadsworth Strathclyde Institute of Pharmacy and Biomedical Sciences University of Strathclyde Glasgow, United Kingdom